The $\mathrm{LGE}_{2}$ skeleton was assembled as outlined in Scheme II. Alkylation of (diethylphosphono)acetone (8) with methyl 7-bromohept- $5(Z)$-enoate ${ }^{12}$ affords 5 whose sodium salt reacts with isopropylidene-L-glyceraldehyde (6) to produce the isomeric enones $4-Z$ and $4-E(1: 2.3)$. Reaction of either isomeric enone with vinyl cuprate $7,{ }^{13}$ prepared from 1 -iodo- $3(S)$-[(tert-butyldimethyl-silyl)oxy]-1( $E$ )-octene ${ }^{14}$ afforded identical mixtures of $9-R R^{15}$ ( $70 \%$ ), which has the $\mathrm{C}-8$ configuration of $\mathrm{LGE}_{2}$, and the $\mathrm{C}-8$ epimer $9-S R^{15}(30 \%)$. Interestingly, Michael addition occurred only in the presence of $\mathrm{MgBr}_{2}$, which presumably serves as a Lewis acid catalyst. ${ }^{16}$ That $9-R R$ and $9-S R$ are epimeric at position 8 is evident from the observation that saponification of either ester afforded identical mixtures of epimeric acids $10-R R(70 \%)$ and 10-SR (30\%).

Unexpectedly, treatment of either $9-R R$ or $9-S R$ with aqueous acetic acid followed by sodium periodate afforded the same 13:1 mixture of $\mathrm{LGE}_{2}$ methyl ester ( $11-R R$ ) and the 8 -epi isomer (11-SR). This remarkably high preference for the required $8 R$ configuration contrasts with the 7:3 equilibrium ratio observed for $10-R R$ and $10-S R$. The 1,2 -dihydroxyethyl substituent in the intermediate vicinal diol $\mathbf{3}$ apparently plays a role in the stereoselective epimerization since efficient interception of this intermediate allows conversion of $9-S R$ to mixtures richer in the 11-SR epimer. Thus, treatment of $9-S R$ with aqueous acetic acid in the presence of sodium periodate yielded a $2: 1$ mixture of $11-R R$ and 11-SR. Further discussion of this remarkable stereoselection is

deferred to a full account of this work. The aldehydic ${ }^{1} \mathrm{H}$ NMR resonance for 11-RR in $\mathrm{CDCl}_{3}$ occurs at $\delta 9.47$ as found for the methyl ester of $\mathrm{LGE}_{2}$ derived from $\mathrm{PGH}_{2}$ while the corresponding resonance for 11-SR occurs at $\delta 9.56$. For comparison with $\mathrm{LGE}_{2}$ methyl ester derived from $\mathrm{PGH}_{2}, 8$-epi-9-epi-LGE 2 methyl ester 11-SS was obtained from $9-S S$ prepared by substituting iso-propylidene-D-glyceraldehyde for the $L$ isomer 6 in Scheme II. The ${ }^{1} \mathrm{H}$ and ${ }^{13} \mathrm{C}$ NMR spectra of $11-R R$, 11-SS, and $\mathrm{LGE}_{2}$ methyl ester obtained from $\mathrm{PGH}_{2}$ are almost identical. To facilitate correlation, these keto aldehydes were converted to fluorenylidene derivatives 12 by chemoselective Wittig reaction with 9-fluorenylidenetri- $n$-butylphosphorane. ${ }^{17}$ The adducts 12 show intense UV absorptions ( $\epsilon_{\max } \simeq 40000$; hexane) at both 258 and 229 nm . Furthermore, in contrast with $\operatorname{LGE}_{2}$ (1), analytically pure fluorenylidene derivatives were readily isolated quantitatively by HPLC on partisil. As expected, 12-RR and the fluorenylidene derivative of $\mathrm{LGE}_{2}$ methyl ester derived from $\mathrm{PGH}_{2}$ exhibit identical ORD curves while the curve for 12-SS is virtually a mirror image (Figure 1).

Levuglandin $\mathrm{E}_{2}$ was prepared in good yield from the ketal $10-R R$ (or $10-S R$ ) by treatment with aqueous acetic acid followed by sodium periodate. The ${ }^{1} \mathrm{H}$ NMR spectrum of synthetic $\mathrm{LGE}_{2}$ is identical with that of the more stable levuglandin obtained from PGH ${ }_{2}$ except for tiny absorptions owing to minor impurities in the latter sample which were absent in the synthetic product. The total synthesis now makes $\mathrm{LGE}_{2}$ readily available. The present results also demonstrate that epimerization of $\mathrm{LGE}_{2}$ at positions 8 or 9 is not extensive under the conditions of its solvent-induced formation from $\mathrm{PGH}_{2}$.
(12) (a) Martel, J.; Toromanoff, E. Germn Patent 2121 361, 1971; Chem. Abstr. 1972, 76, 24712d. (b) Corey, E. J.; Sachdev, H. S. J. Am. Chem. Soc. 1973, $95,8483$.
(13) Corey, E. J.; Beames D. J. J. Am. Chem. Soc. 1972, 94, 7210.
(14) Kluge, A. G.; Untch, K. G.; Fried, J. H. J. Am. Chem. Soc. 1972, 94, 7827.
(15) For compounds $9-11$ the designations $R R, S R$, and $S S$ refer to the absolute configurations of the chiral centers at C-8 and C-9, respectively.
(16) Yamamoto, Y.; Yamamoto, S.; Yatagai, H.; Ishihara, Y.; Maruyama, K. J. Org. Chem. 1982, 47, 119 and references cited therein.
(17) Johnson, A. W.; La Count, R. B. Tetrahedron 1969, 9, 130.

Acknowledgment. This research was assisted financially by Grant GM-21249 from the Division of General Medical Sciences of the National Institutes of Health.

Supplementary Material Available: Physical data and purification of compounds 4-6 and 8-12 (12 pages). Ordering information is given on any current masthead page.

## The Nature of Restrictions in the Binding Site of Rhodopsin. A Model Study

Robert S. H. Liu,* Alfred E. Asato, Marlene Denny, and Dennis Mead

Department of Chemistry, 2545 The Mall University of Hawaii, Honolulu, Hawaii 96822 Received July 31, 1984

Continuing the effort of early workers, ${ }^{1}$ the research teams at Hawaii and elsewhere have recently completed the synthesis of all 16 possible geometric isomers of vitamin A. ${ }^{2}$ Among the 14 stable retinal isomers, 10 have been shown to form visual pigment analogues: 11 -cis, 9 -cis, ${ }^{1} 9$-cis, 13 -cis, ${ }^{1.3} 7$-cis, 7 -cis 9 -cis, 7 cis, 13 -cis, 7 -cis, 9 -cis, 13 -cis, ${ }^{4} 7$-cis, 11 -cis, ${ }^{5} 9$-cis, 11 -cis, ${ }^{6}$ and 7 cis, 9 -cis, 11 -cis. ${ }^{7,8}$ The notable exceptions are the all-trans and the 13 -cis isomers, which do not yield stable analogues. However, it is known that after introduced into the binding site, such geometries can be generated as transient forms, i.e., in lumirhodopsin (or Meta-I) ${ }^{9}$ and photo-Meta-II-465. ${ }^{10}$

The failure of these two isomers to form stable pigments has been rationalized on the ground of a longitudinal restriction of the binding site. ${ }^{11,12}$ This explanation is quite satisfactory for the all-trans isomer, which has the longest distance between the center of the cyclohexenyl ring and the carbonyl carbon. ${ }^{11}$ Since the 13 -cis isomer is not uniquely longer than several other isomers, conformational rigidity was also considered important. ${ }^{12}$
An analysis of longitudinal restrictions based on the length of the chromophore overlooks the tetherlike function of the $n$-butyl group of Lys-296. ${ }^{13}$ Also, the model implies freedom of in-plane motion of the imino carbon relative to a rigid cyclohexenyl ring. This is only partially correct because within the binding site the relative distance of the primary (the protonated Schiff base) and the secondary binding sites (the hydrophobic pocket) is likely to vary only within the limits allowed by conformational changes of the butyl group and to a smaller extent the chromophore itself. We now present a different approach in analyzing the shape of the binding site on the basis of its behavior toward retinal isomers.

[^0]

Figure 1. Molecular model of the tethered chromophore of rhodopsin. The butyl group and the polyene chromophore are in the most relaxed conformation. The distance between the $\alpha$-carbon of Lys-296 and the center of the cyclohexenyl ring is affixed by means of a loose bolt through the $\alpha$-carbon and a Lucite cylinder in the middle of the ring. The Lucite plates are taped to the supporting surface. In this way, only pivotal motions about both centers are permissible. Structures for the remaining 15 isomers were built from this model.

This analysis provides new information related to two-dimensional restrictions of the binding site.

In Figure 1 is shown a model of the retinyl chromophore of rhodopsin bonded to the butyl group of Lys-296. The distance between the $\alpha$-carbon of Lys-296 and the center of the cyclohexenyl ring is affixed in a manner as shown. For clarity only single bonds are shown for the middle portion of the polyene chain.

Within this longitudinal restriction between the primary and secondary binding sites, the model is converted to the remaining 15 geometric isomers of the chromophore. In all structures, the more stable s-trans conformation of all single bonds, other than that of the 6,7 -bond, is maintained. Such a conformation for the most flexible 12,13-bond has been rigorously established for rhodopsin. ${ }^{14}$ On the basis of X-ray crystallographic data a larger ring-chain dihedral angle and a slightly twisted 8,9 -bond are assumed for all 7 -cis isomers. ${ }^{12}$ The model shows that in spite of varying lengths of the isomeric chromophores, ${ }^{11}$ all 16 isomers can be constructed within the given limits. However, there is substantial lateral displacement of atoms and change of conformation of the butyl group. This lateral displacement is largely limited to the area defined by the plane of the chromophore. This is the same plane surrounded by and parallel to the axes of the $\alpha$-helices of the protein, ${ }^{15}$ a relatively open space. Therefore, not surprisingly, so many geometric isomers with a diverse molecular shape form pigment analogues.

The inactivity of 13 -cis- and all-trans-retinal must then be due to factors other than longitudinal restrictions. When the relative positions of the carbon atoms of the two isomeric chromophores are compared with those of all other stable chromophores of active isomers, a possible reason for their failure emerges. Figure 2 is a carbon population map for all 10 active isomers. The van der Waal radii of the outer carbons of this collection therefore define the maximum perimeter of the binding site of rhodopsin. The corresponding positions of the inactive 13 -cis and all-trans isomers are marked separately. The map reveals that only in the shaded area, atoms of the two inactive isomers commonly occupy an area different from those of others. An obvious conclusion is that this region is forbidden to the chromophore, most likely being already occupied by a group protruding from the protein surface, thus providing steric inhibition for the observed negative results. One might further infer from the location of the region and its size, being sufficiently large to contain a three-atom fragment, that this protruding group is a carboxylate anion, i.e., the counterion

[^1]

Figure 2. A two-dimensional map of the binding site of opsin: overlay (a population map) of carbon atoms of all 10 active isomers (see text) of the tethered chromophore of rhodospin (solid dots). Superimposed are carbon positions of the inactive isomers: 13-cis (triangles) and all-trans (circles). The dashed straight lines are the carbon framework of the 11 -cis isomer. The maximum perimeter of the binding site is defined by the van der Waal radii of the outer carbons of the active isomers. The shaded area is that commonly occupied by only the inactive isomers. The dashed circles represent a hypothetical carboxylate anion, which should point perpendicularly to the chromophore. ${ }^{16}$

Table I. Properties of 9-Cis and 11-Cis Isomers of 12-Substituted Pigment Analogues

| substituent isomer | retinal absorptn, $\lambda_{\text {max }}, \mathrm{nm}$ | pigment absorptn, $\lambda_{\text {max }}(\operatorname{det})^{a}$ | pig. yield |
| :---: | :---: | :---: | :---: |
| 12-H, 11-cis ${ }^{\text {b }}$ | 365 | 498 (D) | 100\% |
| $9-\mathrm{cis}^{\text {b }}$ | 363 | 483 (D) | >90\% |
| $12-\mathrm{CH}_{3}, 11$-cis | 290 | 489 (D) | 2\% |
|  |  | 495 (C) | 5\% |
| $9-\mathrm{cis}$ | 359 | 486 (D) | 46\% |
|  |  | 487 (C) | 78\% |
| 12-Cl, 11-cis | 290 | (D) | <2\% |
| 9 -cis | 370 | 488 (D) | ~50\% |
| 12-F, 11-cis ${ }^{\text {c }}$ | 363 | 507 (D) | >90\% |
| $9-\mathrm{cis}^{\text {c }}$ | 362 | 493 (D) | >80\% |

${ }^{a}$ Detergent: digitonin (D) or CHAPS (C). ${ }^{b}$ Data from ref $2,8$. ${ }^{c}$ Data from ref 18.
of the protonated Schiff base. Following the discussion of Hargrave, one is tempted to suggest that this group is Asp-83. ${ }^{136}$ Furthermore, the bidentate nature of the charged group might even qualify it as the "secondary" point charge. ${ }^{16}$

Results from several retinal analogue studies are in support of the conclusion of the location of the binding site restriction. For example, for both 12 -methyl- and 12 -chlororetinal ${ }^{17}$ the surprising results of low yields of pigments from the two 11 -cis isomers while relatively high yields of stable pigments from the corresponding 9 -cis isomers (Table I) can be readily accounted for by the positions of the substituents relative to the forbidden zone. Hence, with the smaller 12 -fluororetinal, both isomers form pigment analogues in high yields. ${ }^{18}$ Failure of pigment formation from a ring-fused retinal analogue ${ }^{19}$ is also in agreement with the above conclusion.

[^2]In summary, this simple analysis allows one to expand beyond the one-dimensional limitation of the earlier model. The conclusions are consistent with the current knowledge of the binding site. Furthermore, the model successfully explains new, unusual analogue results. Continued effort will be directed toward a better understanding of restrictions in the third dimension of the binding site.

Acknowledgment. The work is supported by a grant from the U.S. Public Health Services (AM-17806).

Note Added in Proof. The pivotal motion of the $\alpha, \beta$-bond about the $\mathrm{C}_{\alpha}$ atom probably should be more restricted than what is permitted by the current model, correctly suggested to us by Prof. R. Birge. However, models show that such a refinement has no effect on the conclusions reached in the current study.

Supplementary Material Available: Photographs of molecular models of all remaining 15 isomers of the confined butyl retinylidene chromophore and ${ }^{1} \mathrm{H}$ NMR data ( $\delta, J$, and NOE) of isomers of 12 -methyl- and 12-chlororetinal ( 3 pages). Ordering information is given on any current masthead page.

## Control of Olefination Stereochemistry Using Long-Chain Zirconium Alkylidene Analogues of the "Tebbe" Reagent

Susan M. Clift and Jeffrey Schwartz*
Department of Chemistry, Princeton University Princeton, New Jersey 08544
Received September 10, 1984
The Tebbe compound, ${ }^{1} \mathrm{Cp}_{2} \mathrm{Ti}\left(\mathrm{CH}_{2}\right) \mathrm{ClAlMe}_{2}$, has been shown to be a reagent of general synthetic utility for methylene group transfer to a range of organic carbonyls. ${ }^{2}$ In these reactions, since only $\mathrm{CH}_{2}$ is transferred, the problem of controlling stereochemistry of olefination does not exist. Using long-chain alkylidene analogues ${ }^{3,4}$ of the Tebbe reagent, however, raises this as an important issue. Observations concerning "Wittig-type" reactivity of the Tebbe reagent or its analogues suggested a stepwise olefination process involving an intermediary metallooxetane which then fragments to give the metal oxo complex and the olefinic product. ${ }^{5}$ It could be that stereochemical factors leading to the preferential formation of either possible metallooxetane from an unsymme-trically-substituted metal carbene complex control the product distribution of olefins thus formed. The oxygen atom in the metallooxetane may not provide much differentiating stereochemical information and, therefore, may not be of much use in controlling formation of either metallooxetane or ultimate product distribution. Replacing $(=\mathrm{O})$ with a $(=\mathrm{NR})$ group, however, might provide one method to control stereochemistry in an olefination sequence: the imine, which contains a sterically significant group on nitrogen, would give rise to an N -substituted metallazetidine structure, and the ring that is preferentially formed may be the one that minimizes steric interactions among all four groups of the four-membered ring. With this concept in mind, we have studied reactions between substituted zirconium carbene complexes and imines and find that not only does olefination occur but also that a correlation exists between the $E / Z$ selectivity of olefins formed and the size of the substituent group on imine nitrogen.
(1) Tebbe, F. N.; Parshall, G. W.; Reddy, G. S. J. Am. Chem. Soc. 1978, 100, 3611.
(2) Pine, S. H.; Zahler, R.; Evans, D. A.; Grubbs, R. H. J. Am. Chem. Soc. 1980, 102, 3270.
(3) Hartner, F. W., Jr.; Schwartz, J. J. Am. Chem. Soc. 1981, 103, 4979.
(4) Hartner, F. W., Jr.; Schwartz, J.; Clift, S. M. J. Am. Chem. Soc. 1983, 105, 640 .
(5) Kauffmann, T.; Ennen, B.; Sander, J.; Wieschollek, R. Angew. Chem., Int. Ed. Engl. 1983, 22, 244.

Table I

|  <br> 1a. R=tert$\mathrm{L}=\mathrm{PPh}_{3}$ <br> 16. $\begin{aligned} & R=n-C_{4} t \\ & L=P M e_{2} P h \end{aligned}$ |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: |
| carbene | X | " ${ }^{\prime \prime}{ }^{\circ}$ | $E / Z$ | $\begin{gathered} \mathrm{h} \text { at } \\ 75^{\circ} \mathrm{C} \end{gathered}$ | olefin yield, $\%^{a, b}$ |
| 1 a | $=0$ |  | 0.50 | 3 | 76 |
|  | $=\mathrm{NCH}_{3}$ | 1.8 | 2.06 | 3 | 88 |
|  | $=\mathrm{NCH}\left(\mathrm{CH}_{3}\right)_{2}$ | 2.1 | 2.30 | 4 | 47 |
|  | $=\mathrm{NPh}$ | 3.1 | 2.76 | 3 | 56 |
|  | $=\mathrm{NC}\left(\mathrm{CH}_{3}\right)_{3}$ | $>4.5$ | 4.32 | 2 | 16 |
| 1b | $=0$ |  | 1.11 | 5 | 72 |
|  | $=\mathrm{NCH}_{3}$ | 1.8 | 1.13 | 5 | 85 |
|  | $=\mathrm{NCH}\left(\mathrm{CH}_{3}\right)_{2}$ | 2.1 | 1.67 | 5 | 39 |

${ }^{a}$ Based on carbene; product ratios were determined by VPC analysis; products were isolated by preparative VPC and structures were confirmed by comparison with authentic materials. ${ }^{b} 1$ slowly dimerizes under these conditions; ${ }^{4}$ no carbene complex remained; the unreacted imine can be recovered unchanged. ${ }^{\text {c }}$ For substituents on $\mathrm{N} .{ }^{12}$

## Table II



| carbene | X | $E / Z$ | enol ether <br> yield, $\%^{a}$ |
| :---: | :--- | :---: | :---: |
| $\mathbf{1 a}$ | $=\mathrm{O}$ | 1.00 | 71 |
|  | $=\mathrm{NPh}$ | 0.53 | 72 |
| $\mathbf{1 b}$ | $=\mathrm{O}$ | 1.04 | 80 |
|  | $=\mathrm{NPh}$ | 0.35 | 91 |

${ }^{a}$ Based on carbene complex, determined by VPC. ${ }^{b} 1: 2.3$ ratio.
In a typical olefination procedure benzaldehyde ( 1 equiv) was added to a toluene solution of $1 \mathbf{a}^{4}(0.062 \mathrm{M})$ at $25^{\circ} \mathrm{C}$; the reaction mixture was then heated to $75^{\circ} \mathrm{C}$ for 3 h . A preference for formation of the $Z$ isomer of the olefin was noted as it is in "salt-free" Wittig reactions ${ }^{6}$ using triphenylphosphoranes. ${ }^{6 b}$ In contrast, using the imines shown in Table I this $E / Z$ selectivity can be reversed. ${ }^{7}$ In each case, procedures were identical with the one noted for benzaldehyde. For example, $N$-methylbenzylideneimine ( 1 equiv) was added to a toluene solution of $1 \mathbf{1 a}^{4}$ $(0.062 \mathrm{M})$ at $25^{\circ} \mathrm{C}$, followed by heating to $75^{\circ} \mathrm{C}$ for 3 h . A preference for the $E$ isomer of the olefin was noted.
The Tebbe reagent is especially useful in that it can methylenate esters to give enol ethers. ${ }^{2}$ We had noted that zirconium carbene complexes likewise react readily with esters. ${ }^{4}$ Consistent with the observations described for imines, we observe that imidates ${ }^{8}$ also react with zirconium carbene complexes to yield enol ethers. Here, as in the case of the imines described above, by use of an N substituted imidate, $E / Z$ selectivity of the olefin formed can be controlled (see Table II).

Reactions discovered between zirconium carbene complexes and imines or imidates (and thioketones) expand in scope the nature of substrates that can react with group 4 carbene complexes to give olefins. The fact that a sterically significant group can be utilized in conjunction with a "throwaway" part of the substrate

[^3]
[^0]:    (1) Wald, G.; Brown, P. K.; Hubbard, R. Oroshnik, W. Proc. Natl. Acad. Sci. U.S.A. 1955, 41, 438-450; 1956, 42, 578-580.
    (2) Summarized in: Liu, R. S. H.; Asato, A. E. Tetrahedron 1984, 40, 1931-1969.
    (3) Crouch, R.; Purvin, V.; Nakanishi, K.; Ebrey, T. Proc. Natl. Acad. Sci. U.S.A. 1975, 72, 1538-1542.
    (4) DeGrip, W. J.; Liu, R. S. H.; Ramamurthy, V.; Asato, A. E. Nature (London) 1976, 262, 416-418.
    (5) Kini, A.; Matsumoto, H.; Liu, R. S. H. J. Am. Chem. Soc. 1979, 101, 5978-5079.
    (6) Kini, A.; Matsumoto, H.; Liu, R. S. H. Bioorg. Chem. 1980, 9, 406-410.
    (7) Asato, A. E.; Kini, A.; Denny, M.; Liu, R. S. H. J. Am. Chem. Soc. 1983, 105, 2923-2924.
    (8) Liu, R. S. H.; Matsumoto, H.; Kini, A.; Asato, A. E.; Denny, M.; Kropf, A.; DeGrip, W. J. Tetrahedron 1984, 40, 473-482.
    (9) Yoshizawa, T.; Wald, G. Nature (London) 1963, 197, 1279-1286. For the structure of bathorhodopsin see: Liu, R. S. H.; Asato, A. E. Proc. Natl. Acad. Sci. U.S.A., in press.
    (10) Matthews, R. G.; Hubbard, R.; Brown, P. K.; Wald, G. J. Gen. Physiol. 1963, 47, 215-240.
    (11) Matsumoto, H.; Yoshizawa, T. Vision Res. 1978, 18, 607-609.
    (12) Matsumoto, H.; Liu, R. S. H.; Simmons, C.; Seff, K. J. Am. Chem. Soc. 1980, 102, 4259-4262.
    (13) (a) Ovchinnikov, Y. A.; Abdulaev, N. G.; Feigina, M. Y.; Artamonov, I. D.; Zolotarev, A. S.; Kostina, M. B.; Bogachuk, A. J.; Moroshnikov, A. I.; Martinov, V. I.; Kudelin, A. B. Bioorg. Khim. 1982, 8, 1011-1014. (b) Hargrave, P. A.; McDowell, J. H.; Curtis, D. R.; Wang, J. K.; Juszczak, E.; Fong, S.-L.; Rao, J. K. M.; Argos, P. Biophys. Struct. Mech. 1983, 9, 235-244.

[^1]:    (14) (a) Chan, W. K.; Nakanishi, K.; Ebrey, T. G.; Honig, B. J. Am. Chem. Soc. 1974, 96, 3642-3644. (b) Callender, R. H.; Doukas, A.; Crouch, R.; Nakanishi, K. Biochemistry 1976, 15, 1621-1629.
    (15) Dratz, E.; Hargrave, P. A. Trends Biochem. Sci. (Pers. Ed.) 1983, 8, 128-131.

[^2]:    (16) Honig, B.; Dinur, U.; Nakanishi, K.; Balogh-Nair, V.; Gawinowicz, M. A.; Arnaboldi, M.; Motto, M. G. J. Am. Chem. Soc. 1979, 101, 7084-7086.
    (17) Structural assignments of isomers based on spectral data ( ${ }^{1} \mathrm{H}$ NMR and NOE), which are described in detail in the supplementary material.
    (18) Liu, R. S. H.; Matsumoto, H.; Asato, A. E.; Denny, M.; Shichida, Y.; Yoshizawa, T.; Dahlquist, F. W. J. Am. Chem. Soc. 1981, 103, 7195-7201.
    (19) Chandraratna, R. A. S.; Okamura, W. H. Tetrahedron Lett. 1984, 25, 1003-1006. Chandraratna, R. A. S.; Okamura, W. H., paper presented at US-Japan Joint Symposium on Molecular Mechanisms of the Visual Process, Honolulu, Jan 10-14, 1984.

[^3]:    (6) (a) Vedejs, E.; Meier, G. P.; Snoble, K. A. J. J. Am. Chem. Soc. 1981, 103,2823 . (b) Schlosser, M.; Schaub, B. J. Am. Chem. Soc. 1982. 104, 5821.
    (7) Thioketones also react with zirconium carbene complexes to yield olefins. However, the $E / Z$ selectivity for olefin formation is essentially identical with that noted for the carbonyl compound itself.
    (8) Mukaiyama, T.; Sato, K. Bull. Chem. Soc. Jpn. 1963, 36, 99.

